CPVO-TP/023/2 Final
English
Date: 01/12/2005

European Union
Community Plant Variety Office

PROTOCOL FOR DISTINCTNESS, UNIFORMITY AND STABILITY TESTS

Solanum tuberosum L.

POTATO

UPOV Species Code: SOLAN_TUB

Adopted on 01/12/2005

I - SUBJECT OF THE PROTOCOL

The protocol describes the technical procedures to be followed in order to meet the requirements of Council Regulation 2100/94 on Community Plant Variety Rights. The technical procedures have been agreed by the Administrative Council and are based on general UPOV Document TG/1/3 and UPOV Guideline TG/23/6 dated 31/03/2004 for the conduct of tests for Distinctness, Uniformity and Stability. This protocol applies to all vegetatively propagated varieties of *Solanum tuberosum* L.

II - SUBMISSION OF SEED AND OTHER PLANT MATERIAL

1. The Community Plant Variety Office (CPVO) is responsible for informing the applicant of

- the closing date for the receipt of plant material;
- the minimum amount and quality of plant material required;
- the Examination Office to which material is to be sent.

A sub-sample of the material submitted for test will be held in the variety collection of the Examination Office as the definitive sample of the candidate variety.

2. Final dates for receipt of documentation and material by the Examination Office

The final dates for receipt of requests, technical questionnaires and the final date or submission period for plant material will be decided by the CPVO and each Examination Office chosen.

The Examination Office is responsible for immediately acknowledging the receipt of requests for testing, and technical questionnaires. Immediately after the closing date for the receipt of plant material the Examination Office should inform the CPVO if no plant material has been received. However, if unsatisfactory plant material is submitted the CPVO should be informed as soon as possible.

3. Seed potato requirements

Information with respect to closing dates and submission requirements of plant material for the technical examination of varieties can be found on the CPVO web site (www.cpvo.europa.eu) and in the special issue S2 of the Official Gazette of the Office published yearly at the month of August.

Quality of seed potato material:	The minimum requirements should not be less than the standards laid down in Council Directive 2002/56/EC. The plant material should be visibly healthy and sound such that subsequent growth would not be impaired.
Seed potato treatment:	The plant material must not have undergone any treatment unless the CPVO and the Examination Office allow or request such treatment. If it has been treated, full details of the treatment must be given.
Labelling of sample:	- Species - File number of the application allocated by the CPVO - Breeder's reference - Examination reference (if known) - Name of applicant - The phrase "On request of the CPVO".

III - CONDUCT OF TESTS

1. Variety collection

A variety collection will be maintained for the purpose of establishing distinctness of the candidate varieties in test. A variety collection may contain both living material and descriptive information. A variety will be included in a variety collection only if plant material is available to make a technical examination.

Pursuant to Article 7 of Council Regulation No. 2100/94, the basis for a collection should be the following:

- varieties listed or protected at the EU level or at least in one of the EEA Member States;
- varieties protected in other UPOV Member States;
- any other variety in common knowledge.

The composition of the variety collection in each Examination Office depends on the ecological conditions in which the Examination Office is located.

Variety collections will be held under conditions which ensure the long term maintenance of each accession. It is the responsibility of Examination Offices to replace reference material which has deteriorated or become depleted. Replacement material can only be introduced if appropriate tests confirm conformity with the existing reference material. If any difficulties arise for the replacement of reference material Examination Offices must inform the CPVO. If authentic plant material of a variety cannot be supplied to an Examination Office the variety will be removed from the variety collection.

2. Material to be examined

Candidate varieties will be directly compared with other candidates for Community plant variety rights tested at the same Examination Office, and with appropriate varieties in the variety collection. When necessary an Examination Office may also include other candidates and varieties. Examination Offices should therefore make efforts to co-ordinate the work with other offices involved in DUS-testing of potatoes. There should be at least an exchange of technical questionnaires for each candidate variety, and during the test period, Examination Offices should notify each other and the CPVO of candidate varieties which are likely to present problems in establishing distinctness. In order to solve particular problems Examination Offices may exchange plant material.

3. Characteristics to be used

The characteristics to be used in DUS tests and preparation of descriptions shall be those referred to in the table of characteristics. All the characteristics shall be used, providing that observation of a characteristic is not rendered impossible by the expression of any other characteristic, or the expression of a characteristic is prevented by the environmental conditions under which the test is conducted. In the latter case, the CPVO should be informed. In addition the existence of some other regulation e.g. plant health, may make the observation of the characteristic impossible.

The Administrative Council empowers the President, in accordance with Article 23 of Commission Regulation N° 1239/95, to insert additional characteristics and their expressions in respect of a variety.

4. Grouping of varieties

The varieties and candidates to be compared will be divided into groups to facilitate the assessment of distinctness. Characteristics which are suitable for grouping purposes are those which are known from experience not to vary, or to vary only slightly, within a variety and which in their various states of expression are fairly evenly distributed throughout the collection. In the case of continuous grouping characteristics overlapping states of expression between adjacent groups is required to reduce the risks of incorrect allocation of candidates to groups. The characteristics that could be used for grouping are the following (CPVO numbering; G for grouping in the table of characteristics)

a) Lightsprout: proportion of blue in anthocyanin coloration of base (characteristic 4)

b) Flower corolla: intensity of anthocyanin coloration on inner side (characteristic 28)

c) Flower corolla: proportion of blue in anthocyanin coloration on inner side (characteristic 29)

d) Plant: time of maturity (characteristic 31)

e) Tuber: colour of skin (characteristic 34)

5. Trial designs and growing conditions

The minimum duration of tests will normally be two independent growing cycles. Tests will be carried out under conditions ensuring normal growth. The size of the plots will be such that plants or parts of plants may be removed for measuring and counting without prejudice to the observations which must be made up to the end of the growing cycle.

The test design is as follows:

As a minimum, each test should include a total of 60 plants which should be divided between two or more replicates. The assessment of lightsprout characteristics should be carried out on at least 5 lightsprouts.

All observations determined by measurement or counting should be made on 20 plants or parts of 20 plants.

6. Special tests

In accordance with Article 83(3) of Council Regulation No. 2100/94 an applicant may claim either in the Technical Questionnaire or during the test that a candidate has a characteristic which would be helpful in establishing distinctness. If such a claim is made and is supported by reliable technical data, a special test may be undertaken providing that a technically acceptable test procedure can be devised.

Special tests will be undertaken, with the agreement of the President of CPVO, where distinctness is unlikely to be shown using the characters listed in the protocol.

7. Standards for decisions

a) **Distinctness**

A candidate variety will be considered to be distinct if it meets the requirements of Article 7 of Council Regulation No. 2100/94.

Qualitative characteristics:

In the case of characteristics which show discrete discontinuous states of expression, a difference between two varieties is clear if the respective characteristics have expressions which fall into two different states.

Quantitative characteristics:

Characteristics which show a continuous range of expression from one extreme to the other may be either measured or visually observed.

In the case of visually observed characteristics, a difference between two varieties is clear if the expression of the respective characteristics differs by at least the span of one note, taking into account the variability observed within the varieties.

If the significance level or statistical methods proposed are not appropriate the method used should be clearly described.

b) **Uniformity**

A candidate will be considered to be sufficiently uniform if the number of off-type plants does not exceed 2 in 60 plants examined (population standard of 1% and acceptance probability of ≥ 95%). In a sample size of 5 plants no off-type is allowed.

c) **Stability**

A candidate will be considered to be sufficiently stable when there is no evidence to indicate that it lacks uniformity.
Seed potato samples of further submissions included in any test must show the same expression of characteristics as the material originally supplied.

IV - REPORTING OF RESULTS

After each recording season the results will be summarised and reported to the CPVO in the form of a UPOV model interim report in which any problems will be indicated under the headings distinctness, uniformity and stability. Candidates may meet the DUS standards after two growing cycles but in some cases three growing cycles may be required. When tests are completed the results will be sent by the Examination Office to the CPVO in the form of a UPOV model final report.

If it is considered that the candidate complies with the DUS standards, the final report will be accompanied by a variety description in the format recommended by UPOV. If not the reasons for failure and a summary of the test results will be included with the final report.

The CPVO must receive interim reports and final reports by the date agreed between the CPVO and the Examination Office.

Interim reports and final examination reports shall be signed by the responsible member of the staff of the Examination Office and shall expressly acknowledge the exclusive rights of disposal of CPVO.

V - LIAISON WITH THE APPLICANT

If problems arise during the course of the test the CPVO should be informed so that the information can be passed on to the applicant. Subject to prior permanent agreement, the applicant may be directly informed at the same time as the CPVO particularly if a visit to the trial is advisable.

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VI - TABLE OF CHARACTERISTICS TO BE USED IN DUS-TEST AND PREPARATION OF DESCRIPTION

CPVO N°	UPOV N°	Characteristics	Stage,[1] Method	Examples[2]	Note
1.	**1.**	**Lightsprout: size**	VG		
(+)[3]		small		Grata	3
		medium		Diamant	5
		large		Gloria	7
2.	**2.**	**Lightsprout: shape**	VG		
(+)		spherical		Albas	1
		ovoid		Marabel	2
		conical		Bintje	3
		broad cylindrical		Diamant	4
		narrow cylindrical			5
3.	**3.**	**Lightsprout: intensity of anthocyanin coloration of base**	VG		
(+)		absent or very weak		Estima	1
		weak		Santé	3
		medium		Grandifolia	5
		strong		Granola	7
		very strong		Red Duke of York	9
4.	**4.**	**Lightsprout: proportion of blue in anthocyanin coloration of base**	VG		
(+)		absent or low		Desiree	1
		medium		Pamina	2
G		high		Agria	3

1 The optimum stage of development for the assessment of each characteristic is indicated by a number in the fourth column. The stages of development denoted by each number are described in Annex 1.
2 Example varieties are given as an indication, others may be used.
3 See explanations in Annex 1 in 'Explanations and Methods'

CPVO N°	UPOV N°	Characteristics	Stage,[1] Method	Examples[2]	Note
5.	**5.**	**Lightsprout: pubescence of base**	VG		
(+)		absent or very weak		Santé	1
		weak		Diamant	3
		medium		Junior	5
		strong		Duke of York, Rikea	7
		very strong		Carmona	9
6.	**6.**	**Lightsprout: size of tip in relation to base**	VG		
(+)		small		Quinta	3
		medium		King Edward, Ukama	5
		large		Erntestolz	7
7.	**7.**	**Lightsprout: habit of tip**	VG		
(+)		closed		Quinta	1
		intermediate		Rita	3
		open		Diamant	5
8.	**8.**	**Lightsprout: anthocyanin coloration of tip**	VG		
(+)		absent or very weak		Estima	1
		weak		Duke of York	3
		medium		Spunta	5
		strong		Agria	7
		very strong		Red Duke of York	9
9.	**9.**	**Lightsprout: pubescence of tip**	VG		
(+)		absent or very weak			1
		weak		Quinta	3
		medium		Princess	5
		strong		Elles	7
		very strong			9

CPVO N°	UPOV N°	Characteristics	Stage,[1] Method	Examples[2]	Note
10.	**10.**	**Lightsprout: number of root tips**	VG		
(+)		few		Estima, Sarina	3
		medium		Bintje	5
		many		Belladonna	7
11.	**11.**	**Lightsprout: length of lateral shoots**	VG		
(+)		short		Producent	3
		medium		Estima, Princess	5
		long		Spunta	7
12.	**12.**	**Plant: foliage structure**	1		
(+)		stem type	VG	Agria, Estima	1
		intermediate type		Premiere	2
		leaf type		Kennebec	3
13.	**13.**	**Plant: growth habit**	1		
(+)		upright	VG	Quinta	3
		semi-upright		Desiree, Secura	5
		spreading		Gloria	7
14.	**14.**	**Stem: anthocyanin coloration**	1		
(+)		absent or very weak	VG	Estima	1
		weak		Atlantic	3
		medium		Saturna	5
		strong		Desiree	7
		very strong		Red Duke of York	9
15.	**15.**	**Leaf: outline size**	1		
(+)		small	VG	Kingston, Natalie	3
		medium		Grata	5
		large		Kennebec	7

CPVO N°	UPOV N°	Characteristics	Stage,[1] Method	Examples[2]	Note
16.	**16.**	**Leaf: openness**	1		
(+)		closed	VG	Likaria	1
		intermediate		Premiere	3
		open		Grandifolia	5
17.	**17.**	**Leaf: presence of secondary leaflets**	1		
(+)		weak	VG	Solara	3
		medium		Grata	5
		strong		Hercules	7
18.	**18.**	**Leaf: green colour**	1		
(+)		light	VG	Angela	3
		medium		Ulme	5
		dark		Spunta	7
19.	**19.**	**Leaf: anthocyanin coloration on midrib of upper side**	1		
(+)		absent or very weak	VG	Grata	1
		weak		Russet Burbank	3
		medium		Camilla	5
		strong		Felicitas	7
		very strong		Bildtstar, Roseval	9
20.	**21.**	**Second pair of lateral leaflets: width in relation to length**	1		
(+)		narrow	VG	Russet Burbank	3
		medium		Desiree	5
		broad		Agria	7

CPVO N°	UPOV N°	Characteristics	Stage,[1] Method	Examples[2]	Note
21.	**22.**	**Terminal and lateral leaflets: frequency of coalescence**	1		
(+)		absent or very low	VG	Cherie	1
		low		Bildtstar, Premiere	3
		medium		Agria	5
		high		Romano	7
		very high		Riviera	9
22.	**27.**	**Flower bud: anthocyanin coloration**	1		
(+)		absent or very weak	VG	Grata	1
		weak		Panda	3
		medium		Quinta	5
		strong		Ponto	7
		very strong			9
23.	**28.**	**Plant: height**	2		
		very short	VG	Mimi	1
		short		Atica	3
		medium		Leyla	5
		tall		Grata	7
		very tall		Tomba	9
24.	**29.**	**Plant: frequency of flowers**	2 VG		
		absent or very low		Achat, King Edward	1
		low		Walli	3
		medium		Rita	5
		high		Aiko, Agria	7
		very high		Sibu	9

CPVO N°	UPOV N°	Characteristics	Stage,[1] Method	Examples[2]	Note
25.	**30.**	**Inflorescence: size**	2		
(+)		small	VG	Accent	3
		medium		Grata	5
		large		Karakter	7
26.	**31.**	**Inflorescence: anthocyanin coloration on peduncle**	2		
(+)		absent or very weak	VG	Grata	1
		weak		Aiko	3
		medium		Saturna	5
		strong		Desiree	7
		very strong		Alhamra	9
27.	**32.**	**Flower corolla: size**	2		
(+)		very small	VG	Rhona	1
		small		Sommergold	3
		medium		Grata	5
		large		Karida	7
		very large		Rioja, Roseval	9
28.	**33.**	**Flower corolla: intensity of anthocyanin coloration on inner side**	2 VG		
(+)		absent or very weak		Grata	1
		weak		Secura	3
		medium		Ponto	5
		strong		Artana Pomeroy	7
G		very strong			9
29.	**34.**	**Flower corolla: proportion of blue in anthocyanin coloration on inner side**	2 VG		
(+)		absent or low		Granola	1
		medium		Pamina	2
G		high		Rocket	3

CPVO N°	UPOV N°	Characteristics	Stage,[1] Method	Examples[2]	Note
30.	**35.**	**Flower corolla: extent of anthocyanin coloration on inner side**	2 VG		
(+)		absent or very small		Vitelotte Noir	1
		small		Bildtstar, Rosella	3
		medium		Concurrent	5
		large		Panda	7
		very large		Ponto	9
31.	**36.**	**Plant: time of maturity**	3		
(+)		very early	MG	Christa	1
		early		Cilena	3
		medium		Nicola	5
		late		Aula	7
G		very late		Producent	9
32.	**37.**	**Tuber: shape**	4		
(+)		round	VG	Grata	1
		short-oval		Aula	2
		oval		Diamant	3
		long-oval		Linda	4
		long		Spunta	5
		very long		Pompadour	6
33.	**38.**	**Tuber: depth of eyes**	4		
		very shallow	VG	Duke of York, Nadine	1
		shallow		Agria	3
		medium		Erntestolz	5
		deep		Elles	7
		very deep		Vitelotte Noir	9

CPVO N°	UPOV N°	Characteristics	Stage,[1] Method	Examples[2]	Note
34.	**39.**	**Tuber: colour of skin**	4		
		light beige	VG	Nadine	1
		yellow		Agria, Quarta	2
		red		Desiree	3
		red parti-coloured		Cara	4
		blue		Vitelotte Noir	5
		blue parti-coloured		Kestrel, Catriona	6
G		reddish brown		Umatilla Russet	7
35.	**40.**	**Tuber: colour of base of eye**	4		
		white	VG	Nadine	1
		yellow		Agria	2
		red		Quarta	3
		blue		Vitelotte Noir	4
36.	**41.**	**Tuber: colour of flesh**	4		
		white	VG	Russet Burbank	1
		cream		Desiree, Estima	2
		light yellow		Diamant	3
		medium yellow		Bildtstar, Quarta	4
		dark yellow		Princes	5
		red		Red Salad	6
		red parti-coloured		Early Rose	7
		blue		Vitelotte Noir	8
		blue parti-coloured		Herd Laddie	9

CPVO N°	UPOV N°	Characteristics	Stage,[1] Method	Examples[2]	Note
37.	**42.**	**<u>Light beige and yellow skinned varieties only</u>: Tuber: anthocyanin coloration of skin in reaction to light**	4 VG		
(+)		absent or very weak		Estima	1
		weak		Diamant	3
		medium		Charlotte	5
		strong		Granola	7
		very strong			9

ANNEXES TO FOLLOW

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I. ANNEX I

II. ANNEX II

ANNEX I

EXPLANATIONS ON THE TABLE OF CHARACTERISTICS

Method of observation of characteristics

Letters indicate the relevant method of assessment:

VG	Visual assessment by a single observation of a group of plants or plant parts
MG	Measurement of a group of plants or plant parts

Optimal Stage of Development for the Assessment of Characteristics

1 = bud stage
2 = flowering stage
3 = ripening stage of tubers
4 = after harvest

Unless otherwise stated, all observations with respect to certain characteristics should be made on the part of the plants as mentioned hereunder:

Lightsprouts (Characteristics 1 – 11): All observations on the lightsprout should be made on a total of 5 tubers as a minimum according to the following method:

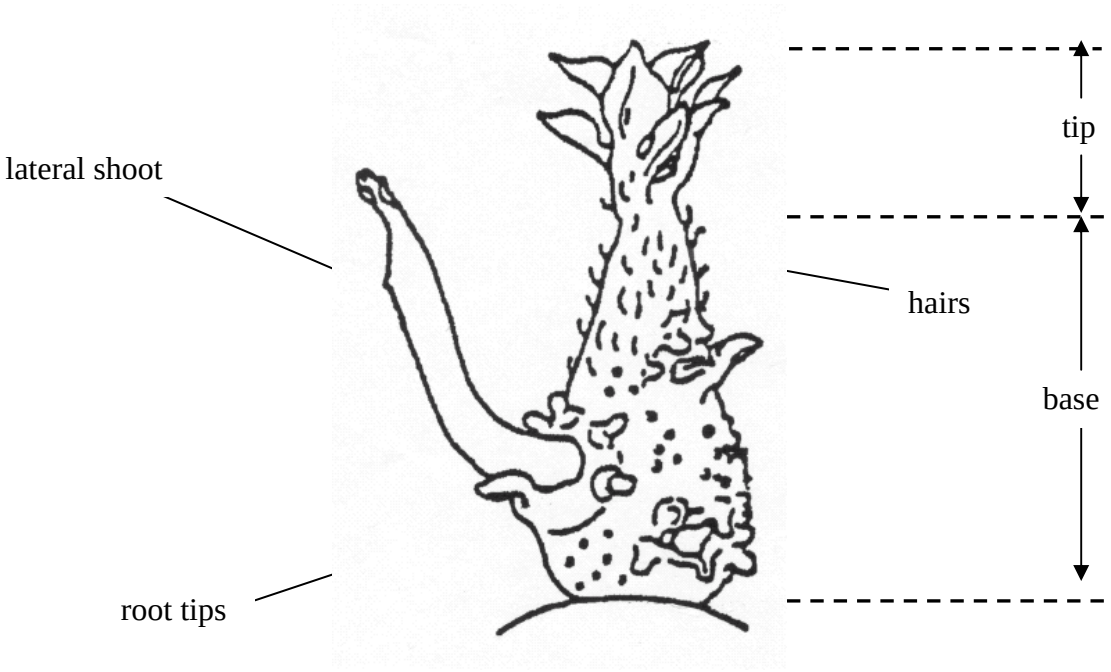
The spectrum and the intensity of the light source are the most important factors for the expression of lightsprout characteristics. This spectrum is defined by the type of lamps and the voltage used. When extremes of temperature are avoided, the influence of the temperature on the speed of development is small. A good expression of characteristics is obtained when the lightsprouts are grown in a light-sealed cabinet at room temperature under continuous light provided by small incandescent bulbs (6V AC/0.05 A) giving an intensity of 5 to 10 lux (approximately 8 bulbs per square meter, 25-40 cm above the tubers).

Leaf (Characteristics 15 – 17; 20): All observations should be made on fully developed leaves from the centre of the plant. One leaf from each of 20 plants should be picked from a main stem midway between the top and the bottom of the plant.

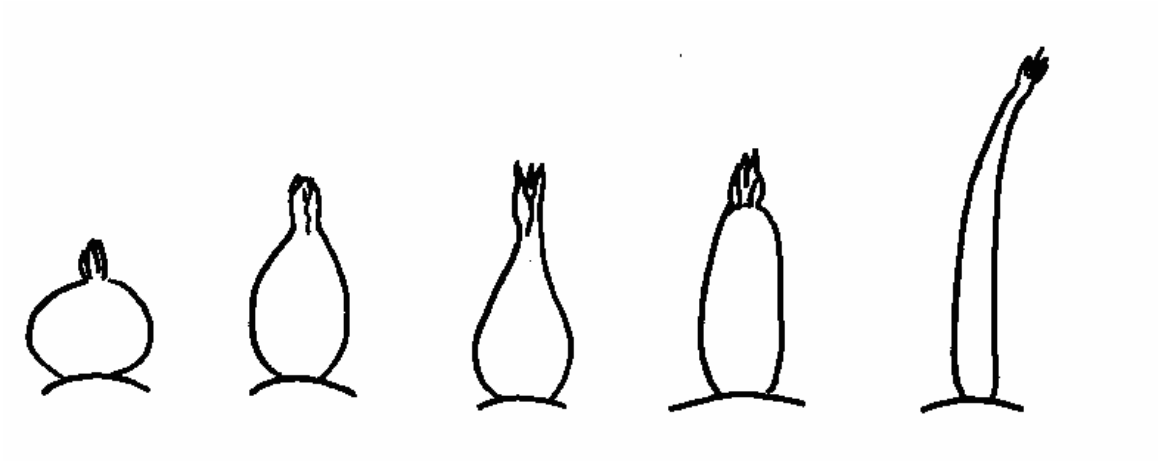
Leaf (Characteristic 18 + 19; 21): All observations on the leaf should be made on fully developed leaves from the centre of the plant.

Flower (Characteristics 27 – 30): All observations of flower colour should be made on the inner side of freshly opened flowers.

Ads. 1 to 11: Lightsprout



Ad. 2: Lightsprout: shape



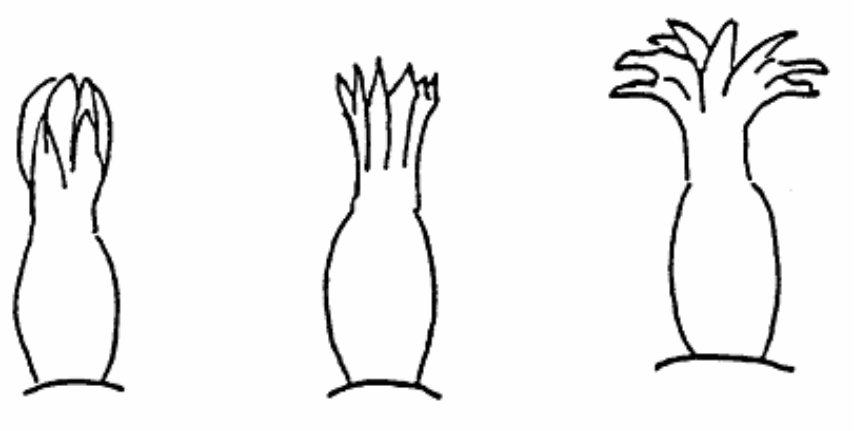
Ad. 3: Lightsprout: intensity of anthocyanin coloration of base

If the intensity of the anthocyanin coloration is “absent”, the lightsprout appears green.

Ads. 4: Lightsprout: proportion of blue in anthocyanin coloration of base,
and 29: Flower corolla: proportion of blue in anthocyanin coloration on inner side

The colour of anthocyanin results from a red and a blue component. If the proportion of blue is low the anthocyanin appears red-violet. If the proportion of blue is high the anthocyanin appears blue-violet.

Ad. 7: Lightsprout: habit of tip



1	3	5
Closed	intermediate	open

The characteristic should be observed after about 10 weeks to obtain a good differentiation in the collection.

Ad. 11: Lightsprout: length of lateral shoots

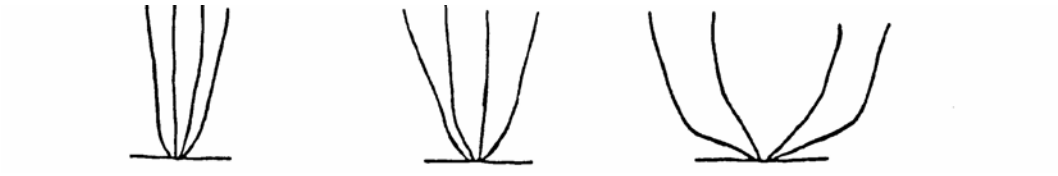
3	5	7
short	medium	long

Ad. 12: Plant: foliage structure

Stem type: foliage open, stems clearly visible
Intermediate type: foliage half open, stems partly visible
Leaf type: foliage closed, stems not, or hardly, visible

1	2	3
stem type	intermediate type	leaf type

Ad. 13: Plant: growth habit



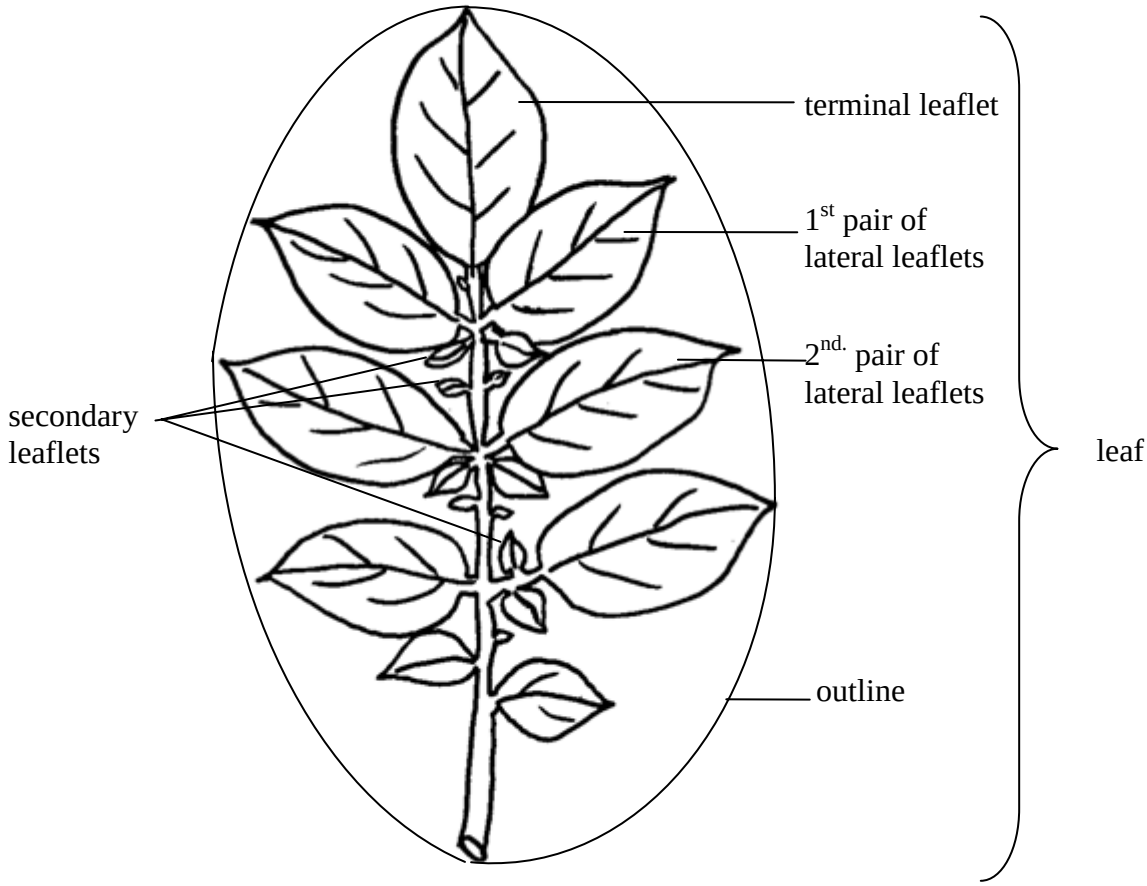
3	5	7
upright	semi-upright	spreading

Ads. 14, 19, 22, 26, 30: Anthocyanin coloration

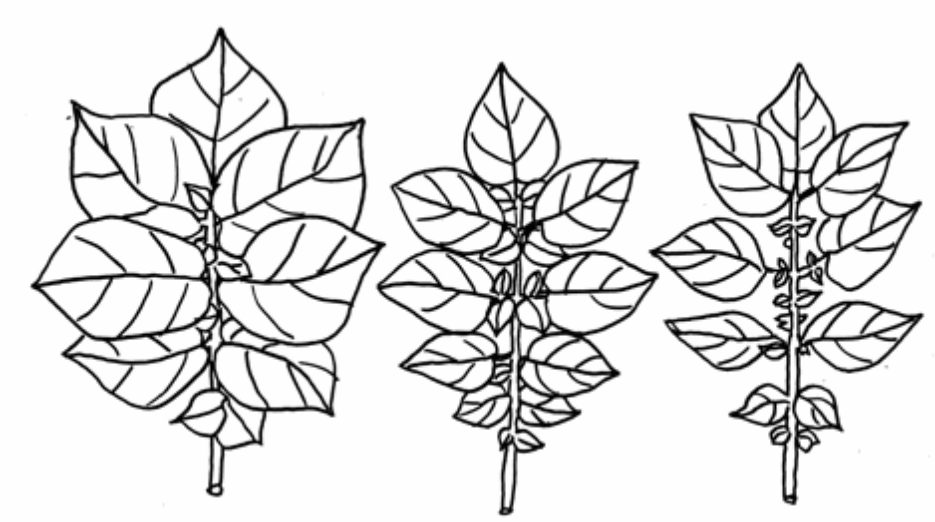
The extent of anthocyanin coloration should be observed in relation to the total area. Distribution and intensity should not be considered.

The extent of anthocyanin coloration of flower buds should be observed on fully developed buds before the corolla is visible.

Ads. 15 to 21: Leaf characteristics

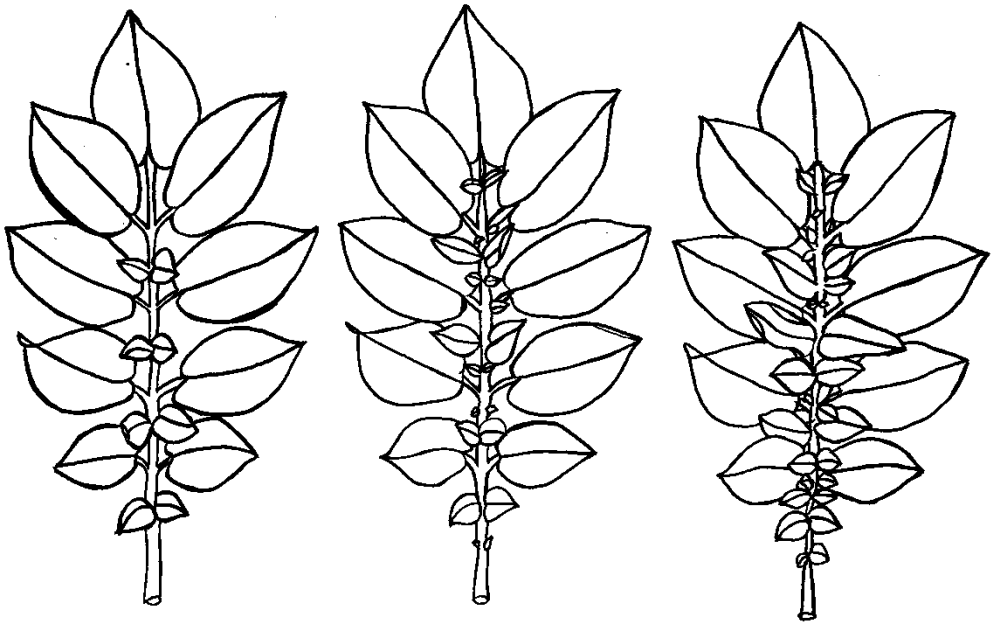


Ad. 16: Leaf: openness



1	3	5
closed	intermediate	open

Ad. 17: Leaf: presence of secondary leaflets

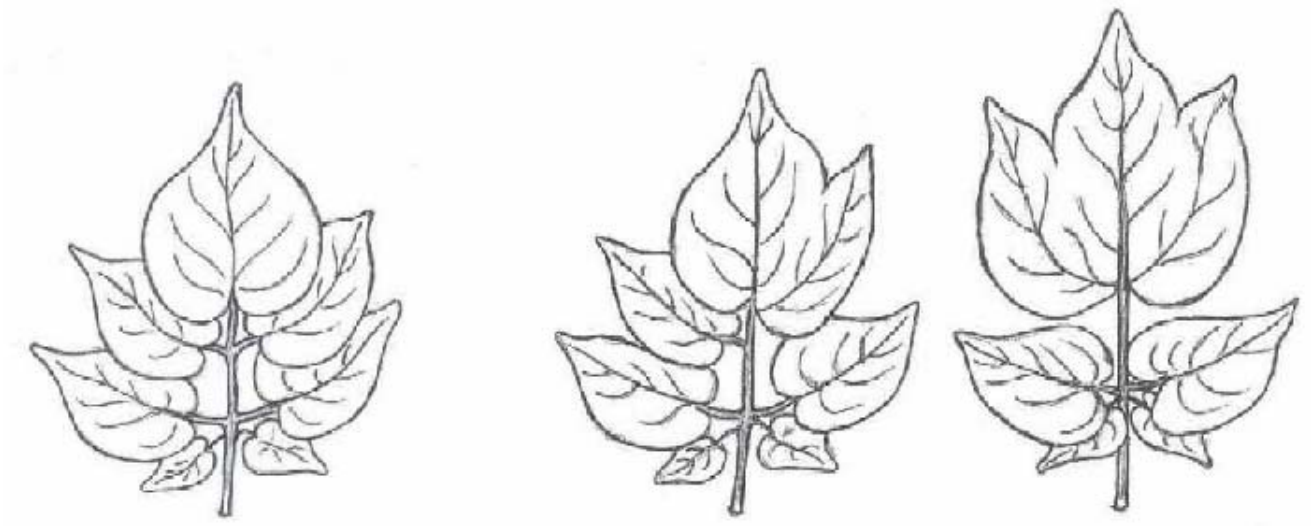


3	5	7
weak	medium	strong

Ad. 20: Second pair of lateral leaflets: width in relation to length

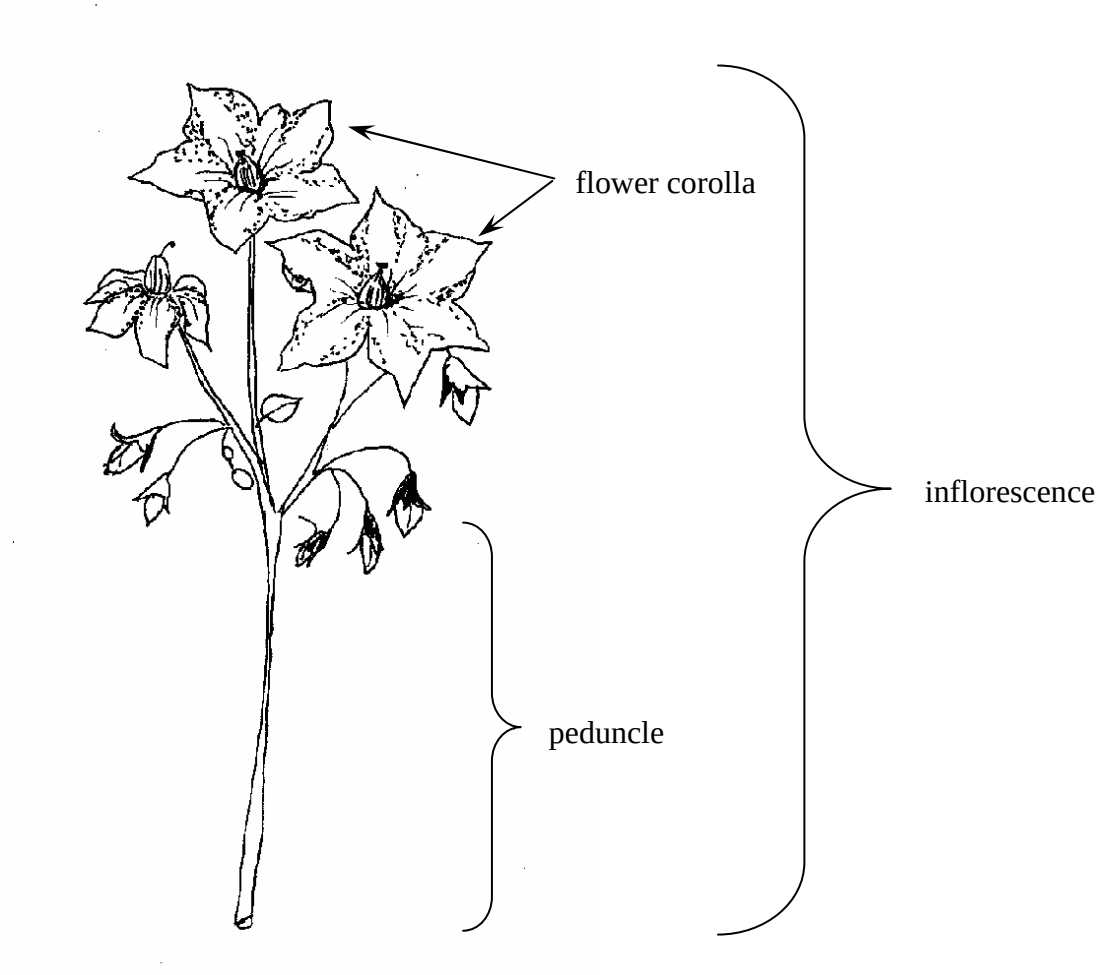
3	5	7
narrow	medium	broad

Ad. 21: Terminal and lateral leaflets: frequency of coalescence



not coalescent coalescent

Ads. 25–30: Inflorescence and flower characteristics



Ad. 28: Flower corolla: intensity of anthocyanin coloration on inner side

If the intensity of the anthocyanin coloration on the inner side is “absent”, the flower corolla appears white.

Ad. 31: Plant: time of maturity

The time of maturity is reached when 80% of the leaves are dead.

Ad. 32: Tuber: shape

1	2	3	4	5	6
round	short-oval	oval	long-oval	long	very long

The predominant shape should be observed on the harvested material from each plot.

Ad. 37: Light beige and yellow skinned varieties only: Tuber: anthocyanin coloration of skin in reaction to light

The anthocyanin development in the skin of light beige and yellow skinned varieties should be assessed after 10 days of exposure to full daylight or after 150 hours of exposure to artificial light.

ANNEX II

European Union
Community Plant Variety Office

TECHNICAL QUESTIONNAIRE

to be completed in connection with an application for Community Plant Variety Rights
Please answer all questions. A question without any answer will lead to a non-attribution of an application date. In cases where a field / question is not applicable, please state so.

1. **Botanical taxon:** Name of the genus, species or sub-species to which the variety belongs and common name

***Solanum tuberosum* L.**

POTATO

2. **Applicant(s):** Name(s) and address(es), phone and fax number(s), Email address, and where appropriate name and address of the procedural representative

3. **Variety denomination**

a) Where appropriate proposal for a variety denomination:

b) Provisional designation (breeder's reference):

4. Information on origin, maintenance and reproduction of the variety

4.1 Origin

(a) Seedling (indicate parent varieties).....[ ]

(b) Mutation (indicate parent variety) .....[ ]

(c) Discovery (indicate where, when and how the variety has been developed).....[ ]

(d) Other (please specify).....[ ]

4.2 Method of propagating the variety

4.2.1 Vegetative propagation

(a) tuber.....[ ]

(b) other (state method).....[ ]

4.2.2 Other .....[ ]
(please provide details)

4.3 Geographical origin of the variety: the region and the country in which the variety was bred or discovered and developed

5. Characteristics of the variety to be indicated (the number in brackets refers to the corresponding characteristic in the CPVO Protocol; please mark the state of expression which best corresponds).

	Characteristics	Example varieties	Note
5.1 (24)	**Plant: frequency of flowers**		
	absent or very low	Achat, King Edward	1 [ ]
	low	Walli	3 [ ]
	medium	Rita	5 [ ]
	high	Aiko, Agria	7 [ ]
	very high	Sibu	9 [ ]
5.2 (28)	**Flower corolla: intensity of anthocyanin coloration of inner side**		
	absent or very weak	Grata	1 [ ]
	weak	Secura	3 [ ]
	medium	Ponto	5 [ ]
	strong	Artana, Pomeroy	7 [ ]
	very strong		9 [ ]
5.3 (29)	**Flower corolla: proportion of blue in anthocyanin coloration of inner side**		
	absent or low	Granola	1 [ ]
	medium	Pamina	2 [ ]
	high	Rocket	3 [ ]
5.4 (31)	**Plant: time of maturity**		
	very early	Christa	1 [ ]
	early	Cilena	3 [ ]
	medium	Nicola	5 [ ]
	late	Aula	7 [ ]
	very late	Producent	9 [ ]

	Characteristics	Example varieties	Note
5.5 (32)	**Tuber: shape**		
	round	Grata	1 [ ]
	short-oval	Aula	2 [ ]
	oval	Diamant	3 [ ]
	long-oval	Linda	4 [ ]
	long	Spunta	5 [ ]
	very long	Pompadour	6 [ ]
5.6 (34)	**Tuber: colour of skin**		
	light beige	Nadine	1 [ ]
	yellow	Agria, Quarta	2 [ ]
	red	Désirée	3 [ ]
	red parti-coloured	Cara	4 [ ]
	blue	Vitelotte Noir	5 [ ]
	blue parti-coloured	Catriona	6 [ ]
	reddish brown	Umatilla Russet	7 [ ]
5.7 (35)	**Tuber: colour of base of eye**		
	white	Nadine	1 [ ]
	yellow	Agria	2 [ ]
	red	Quarta	3 [ ]
	blue	Vitelotte Noir	4 [ ]

Characteristics		Example varieties	Note
5.8 (36)	**Tuber: colour of flesh**		
	white	Russet Burbank	1 [ ]
	cream	Desiree, Estima	2 [ ]
	light yellow	Diamant	3 [ ]
	medium yellow	Bildtstar, Quarta	4 [ ]
	dark yellow	Princess	5 [ ]
	red	Red Salad	6 [ ]
	red parti-coloured	Early Rose	7 [ ]
	blue	Vitelotte Noir	8 [ ]
	blue parti-coloured	Herd Laddie	9 [ ]

6. Similar varieties and differences from these varieties:

Denomination of similar variety	Characteristic in which the similar variety is different[1)]	State of expression of similar variety	State of expression of candidate variety

[1)] In the case of identical states of expressions of both varieties, please indicate the size of the difference

7. Additional information which may help to distinguish the variety

7.1 Resistance to pests and diseases

7.2 Special conditions for the examination of the variety

[ ] YES, please specify

[ ] NO

7.3 Other information

[ ] YES, please specify

[ ] NO

8. GMO-information required

The variety represents a Genetically Modified Organism within the meaning of Article 2(2) of Council Directive EC/2001/18 of 12/03/2001.

[ ] YES [ ] NO

If yes, please add a copy of the written attestation of the responsible authorities stating that a technical examination of the variety under Articles 55 and 56 of the Basic Regulation does not pose risks to the environment according to the norms of the above-mentioned Directive.

I/we hereby declare that to the best of my/our knowledge the information given in this form is complete and correct.

Date Signature Name

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